

REVIEW OF RESEARCH

IMPACT FACTOR : 5.7631(UIF)

ISSN: 2249-894X

VOLUME - 1 | ISSUE - 3 | MARCH - 2019

HUMAN FUNGAL PATHOGENIC DIVERSITY

Jadhav Vyankatesh Abasaheb Assistant professor and Research Scholar, Department of Biophysics, Digambarrao Bindu College(ACS), Bhokar, Dist.Nanded (M.S.)-431801. vbiophysics@gmail.com

ABSTRACT :

Yeast and Molds are the predominate forms of fungal eukaryotic microorganism. The chitin composition of cell wall in kingdom fungi makes organisms distinguished from eukaryotic life kingdoms of animals and plants. As fungi do not photosynthesis, the heterotypic mode of food absorption from environment by secreting the extracellular enzymes outside the cell. These evolutionary relationships generated by comparing a single gene sequence, generally the small subunit ribosomal RNA gene (SSU rRNA). Since then, information from several protein coding genes has revealed diversity and phylogenetic trees of fungus. In present research, elaborating the conserved targets in phylogeny of genus candida with five species for potential drug targets. Hwp1 is significant cell surface protein used as reference have role in C. albicans biofilm formation and thus an opportunistic therapeutic target.

Keywords: Fungi, phylogeny, Candida, conserved sites, drug targets, Hwp1

1.INTRODUCTION

Fungi have ancient origins, with evidence from fossil record indicating their first appearance about one billion years ago [1]. They belong one of most diverse groups of living organisms. The oldest plant fossils found fungal hyphae confirming that fungi are an extremely ancient group. However, regardless of fossil record, biochemical characters and molecular level have served as potential tool in tracing the probable evolutionary relationships of fungi [2].

Fungal groups can be related by presence of chitin in cell wall, composition of amino acid tryptophan in enzymes and synthesis of lysine in biochemical pathways. Later on, Molecular phylogenetic analyses that became possible during the 1990s have greatly contributed to the understanding of fungal origins evolution in their diversity.

Initially, these evolutionary relationships generated by comparing a single gene sequence, generally the small subunit ribosomal RNA gene (*SSU rRNA*). Since then, information from several protein coding genes has revealed diversity and phylogenetic trees of fungi. Now a day, phylogenetic trees built using a wide variety of data and databases. In

present work, we are exploring the conserved sites of five *Candida* species fungal diversity [3]. The five *candida* species are namely, *C. albicans, C. Africana, C. dubliniensis CD36, C. parapsilosis, C. tropcalis.* Hwp1 is significant cell surface protein required for *C. albicans* biofilm formation in vivo [4] and is thus an opportunistic therapeutic target.

2. METHODS AND METHODOLOGY

2.1NCBI-Protein (https://www.ncbi.nlm.nih.gov/protein)

The National center for biotechnology Information facilities the access to genomic information in form of taxonomy, sequences, structure, function etc., the another resource, Protein database is collection

of sequence submitted from different sources. Protein sequences are the fundamental determinants of biological structure to function relationship [5].

2.2NCBI-FASTA Format (https://www.ncbi.nlm.nih.gov/BLAST/fasta.shtml)

A single line FASTA format followed by lines of sequence data. A greater-than (">") symbol at the beginning description line (define) is distinguished from the sequence data by. It is recommended that all lines of text be shorter than 80 characters in length [6].

2.3 Clustal Omega (https://www.ebi.ac.uk/Tools/msa/clustalo/)

A new multiple sequence alignment program that working on seeded guide trees and HMM profileprofile techniques to generate alignments between minimum three **or more** sequences. In case of two sequences pair wise alignment in preferred [7].

2.4 ProtParam (https://web.expasy.org/protparam)

ProtParam tool allows the analysis of various physical and chemical parameters for a given protein stored in Swiss-prot and user protein sequence. The calculation of parameters includes the mol. weight, theoretical pl, amino acid composition, atomic composition and extinction coefficient etc [8].

2.5 Uniprot Knowledgebase (UniprotKB) (https://www.uniprot.org/uniprot)

A comprehensive, high-quality and freely available resource of protein sequence and function. Uniprot Knowledgebase (UniprotKB) facilitates the information to biological Function of protein. [9]

2.6 InterPro (https://www.ebi.ac.uk/interpro/)

InterPro provides families and domains sites for functional analysis of proteins. It produces a powerful integrated database and diagnostic tool [10].

3. RESULTS

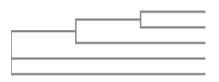
Significant protein Hwp1 in *Candida* species with FASTA Format has following GenBank accession:

Sr.No.	Candida species	GenBank ID				
1	Candida albicans	>ACN63125.1				
2	Candida Africana	>ACB11588.1				
3	Candida dubliniensis CD36	>XP_002419994.1				
4	Candida parapsilosis	>AQY56716.1				
5	Candida tropicalis	>ASK40159.1				
Common NCDL FACTA						

Table1: Hyphal wall protien1 GenBank ID

Source: NCBI-FASTA <u>www.ncbi.nlm.nih.gov/BLAST/fasta.shtml</u>

Clastal omega results for job clustalo-I20190227-134132-0491-99665060-p1m are in form of phylogenetic tree as shown below:



AQY56716.1 -0.05719 ASK40159.1 0.05719 ACN63125.1 -0.0429 XP_002419994.1 0.20334 ACB11588.1 0.03579

Fig 1: Phylogenetic tree for five *candida* species using Neighbour-joining method.

"Advances in Fisheries, Biological and Allied Research"

The phylogenetic tree reveals the *C. tropicalis and C. parapsilosis* more similar species with each other and *C. Africana. Whereas, C. albicans* and *C. Africana* are more distinct among the comparative species [Fig. 1].

Function of Hyphal wall protien1 (Gene name: Hwp1):

- Plays a role in adhesion and is required for mating, normal hyphal development, and cell-to-cell contact [10].
- ✓ It is also revealed functions necessity in biofilm integrity. The adherence to host and participation in virulence. It promotes effective interactions in colonization [11].
- ✓ Hyphal wall protien1 plays a crucial role in gastrointestinal, mucosal, asymptomatic infections and candidacies, caused by the combined action of fungal virulence factors and host inflammatory responses in immunocompromised individuals.

The physicochemical parameters of hyphal wall protein1 has average 1533 Amino Acids in *Candida* Species having Theoretical pl 7.55 highest in *Candida parapsilosis* with average value. Extinction coefficients (assuming all pairs of Cys residues form cystines) are 3184.4. The Average Estimated half and Instability index is 18hrs and 64.99 the Aliphatic Index has highest value for *Candida parapsilosis* (Table. 2).

Parameters/ C.Species	рІ	Aa	Ext. coeff	half-life (hrs.)	Insta bility Index	Aliphatic Index
Ca	3.93	264	4845	1.9	68.61	54.70
Caf	3.61	558	3137	30	72.93	53.33
Cd	3.90	421	3074	30	61.34	66.70
Ср	7.55	236	4220	30	39.98	73.56
Ct	3.56	269	3230	1	82.11	57.03

Table-2: Computation of physicochemical properties of Hwp1 Proteins of Five Candida species.

Domain and repeats:

The conserved and functional 'non-cytoplasmic domain' region of 20-263 aa and four signal peptides at different sits were observed in InterPro results.

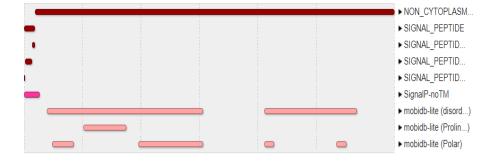


Fig 2: Domain and repeats of hyphal wall protien1 of C. albicans (strain SC5314 / ATCC MYA-2876).

4. CONCLUSION

Fungi have ancient origins; belong to one of most diverse groups of living organisms. Presently, the biochemical characters and molecular level have served as potential tool in tracing evolutionary relationships of fungi.

Source: https://web.expasy.org/cgi-bin/protparam/protparam

[&]quot;Advances in Fisheries, Biological and Allied Research"

Review of Research

These evolutionary relationships generated by comparing a single and multiple gene as well as protein sequence. Since then, information from several protein coding genes has revealed diversity and phylogenetic trees of genus *candida*. In our study five species of *Candida* genus were studied with conserved targets in phylogeny for potential drug targets.

Results are showing *candida* species are having similarity with two distinct species. The physicochemical properties such as half life, stability and aliphatic index signify relevant information about survival and functional environment of protein. Amino acid composition (Positive and Negative) and theoretical pl are related to eachother except *C. parapolis*. The

InterPro result shows region of domain and repeats of non-cytoplasmic domain and four sites of signal peptide.

Thus, Hwp1 is significant cell surface protein required for *C. albicans* biofilm formation was considered as opportunistic therapeutic target.

5. ACKNOWLEDGEMENT

Here, I am very much thankful to my Research Guide and Lab mates for their continues support in research.

6. REFERENCES

- 1. Bonfante, P., & Genre, A. (2008). Plants and arbuscular mycorrhizal fungi: an evolutionarydevelopmental perspective. *Trends in plant science*, *13*(9), 492-498.
- 2. Smith, S. E., & Read, D. J. (2010). Mycorrhizal symbiosis. Academic press.
- 3. McLaughlin, D. J., Hibbett, D. S., Lutzoni, F., Spatafora, J. W., & Vilgalys, R. (2009). The search for the fungal tree of life. *Trends in microbiology*, *17*(11), 488-497.
- 4. Nobile, C. J., Nett, J. E., Andes, D. R., & Mitchell, A. P. (2006). Function of *Candida* albicans adhesin Hwp1 in biofilm formation. *Eukaryotic cell*, *5*(10), 1604-1610.
- Sayers, E. W., Agarwala, R., Bolton, E. E., Brister, J. R., Canese, K., Clark, K., ... & Holmes, J. B. (2019). Database resources of the National Center for Biotechnology Information. *Nucleic acids research*, 47(Database issue), D23
- 6. Booher, N. J., & Bogdanove, A. J. (2014). Tools for TAL effector design and target prediction. *Methods*, 69(2), 121-127.
- Sievers, F., Wilm, A., Dineen, D., Gibson, T. J., Karplus, K., Li, W., ... & Thompson, J. D. (2011). Fast, scalable generation of high-quality protein multiple sequence alignments using Clustal Omega. *Molecular systems biology*, 7(1), 539.
- 8. Walker, J. M. (Ed.). (2005). *The proteomics protocols handbook*. Humana Press.
- Apweiler, R., Attwood, T. K., Bairoch, A., Bateman, A., Birney, E., Biswas, M., ... & Durbin, R. (2001). The InterPro database, an integrated documentation resource for protein families, domains and functional sites. *Nucleic acids research*, 29(1), 37-40.
- 10. Gow, N. A., Van De Veerdonk, F. L., Brown, A. J., & Netea, M. G. (2012). *Candida albicans* morphogenesis and host defence: discriminating invasion from colonization. *Nature reviews microbiology*, *10*(2), 112.
- 11. Mavor, A. L. (2004). *Transcriptional regulation of morphogenesis in Candida albicans* (Doctoral dissertation, University of Aberdeen (United Kingdom).